

## Protocol for Labelling EV Samples with AcoDyes<sup>™</sup> for

## **Detection on Plate Reader**

Packaging	Number of Tests	Volume of Solvent to Add	Solvent	Stock Concentration	Storage Conditions
0.5mL Vials	25 Tests (Trial Size)	ests (Trial Size) 25µL Aqueous			2 to 8%C
	100 Tests	100µL	Buffer	zομΜ	21080

### Table 1. Volume of solvent to add to obtain 25µM stock solution of AcoDyes<sup>™</sup>.

- 1. To get a 25µM stock solution, reconstitute the dried dye in an aqueous buffer of your choice (e.g. sterile, filtered PBS). Refer to Table 1 above for the volume of solvent to add into the dye vial based on the product size (number of tests).
- 2. Vortex the dye for 1 minute to fully dissolve the solid.
- 3. Sonicate the dye for 15 minutes at 40°C.
- 4. Dyes perform best when freshly reconstituted. Solubilised dyes can be stored for up to 1 month at 4°C, but should be vortexed and sonicated for 15 minutes at 40°C before use.
- 5. **[Only for Aco-430<sup>™</sup>, Aco-490<sup>™</sup>, Aco-600<sup>™</sup>]** Aliquot a suitable volume of 25µM stock solution into a clean Eppendorf tube and dilute to 10µM using the same aqueous buffer.
- 6. Dilute EV sample to approximately **10<sup>10</sup> particles/mL** using the same aqueous buffer.
- 7. In a 96-well clear polystyrene non-tissue culture treated plate, prepare the samples according to Table 2 below, mixing thoroughly. Avoid using the outer wells of the plate.

\*AcoDyes<sup>TM</sup> are provided in vials of  $25\mu$ M stock concentration post-reconstitution. We recommend optimising staining for your sample by diluting the dye reagent in aqueous buffer and adding directly to your sample (final dye dilutions of 0.5 $\mu$ M, 1 $\mu$ M, 2.5 $\mu$ M, 5 $\mu$ M). Immediately post-addition, 1-2 hours incubation time is recommended at 37°C.



# Table 2. Recommended EV sample preparation protocol. Ensure that the ratio of dye particles to EV particles remains constant if you plan to deviate from the suggested volumes in Table 2.

Samples to Run	Final Dye Conc* (µM)	Dye Solution		Volume	Volume of	
		Conc* (µM)	Volume* (µL)	of PBS* (µL)	10°/mL EV (μL)	Incubation
EV + Aco-430™	0.5	10	5	5	90	37°C for 1 hour, protected from light
EV + Aco-490 <sup>™</sup>	0.5	10	5	5		
EV + Aco-600™	1.0	10	10	-		
EV + Aco-800™	5.0	25	20	-	80	37°C for 2 hours, protected from light
EV Only	-	-	-	10-20	80-90	Depends on AcoDye™
PBS Only	-	-	-	100	-	

8. Add to plate reader and collect full fluorescence spectrum or fluorescence at peak maxima.



**Figure 1.** PC3 Exosomes (Abcam) at approximately  $10^9$  particles/mL were mixed with  $1\mu$ M of (left) Aco-430, (centre) Aco-490 or (right) Aco-600. Samples were incubated for 1 hour at 37°C before measuring on the Tecan Spark Microplate Reader. The samples were excited at their maximum absorption peak wavelength (refer to Table 3 below) and the full fluorescence spectra were collected.



Dye Name	Aco-430	Aco-490	Aco-600	Aco-800
Appearance	Colourless	Yellow	Purple	Green
$\lambda_{abs,  with  SUVs}$ (nm)	369	422	525	735
$\lambda_{em,  with  SUVs}$ (nm)	403 - 460	458 - 508	586 - 635	775 - 818
Suitable Laser(s)	UV (355 nm)	Violet (405 nm)	Blue or Yellow (488 or 561 nm)	Red (638 nm)
Quantum Yield (%)	94	60	27	13.8
Fluorescence Lifetime (ns)	1.1	0.9	1.9	1.6
IC₅₀ (µM)	>256	>256	170	>272

#### Table 3. Photophysical properties of Aco-430<sup>™</sup>, Aco-490<sup>™</sup>, Aco-600<sup>™</sup> and Aco-800<sup>™</sup>.



Figure 2. Absorption and emission spectra for Aco-430<sup>™</sup>, Aco-490<sup>™</sup>, Aco-600<sup>™</sup> and Aco-800<sup>™</sup>.

Please feel free to email hello@acoerela.com for assistance with troubleshooting of the staining protocol.